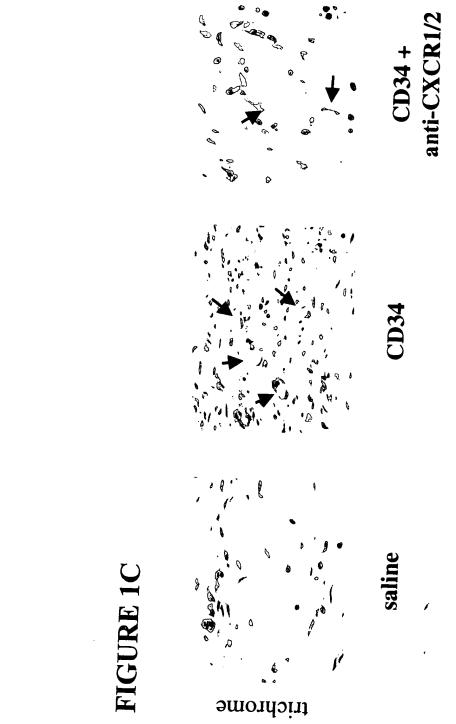


% inhibition of CD34+ angioblasts to ischemic myocardium

FIGURE 1B



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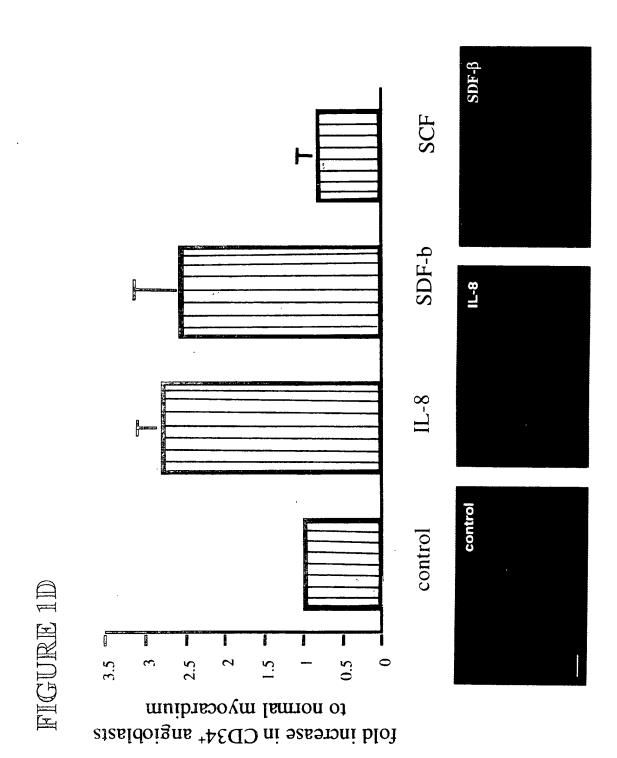
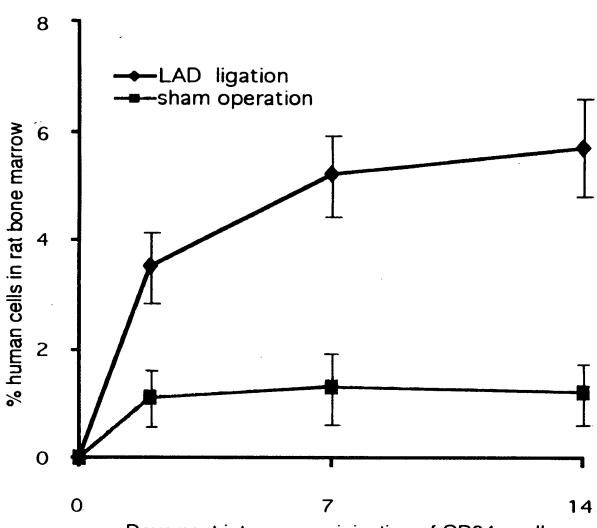


FIGURE 2A



Days post intravenous injection of CD34+ cells

FIGURE 2B

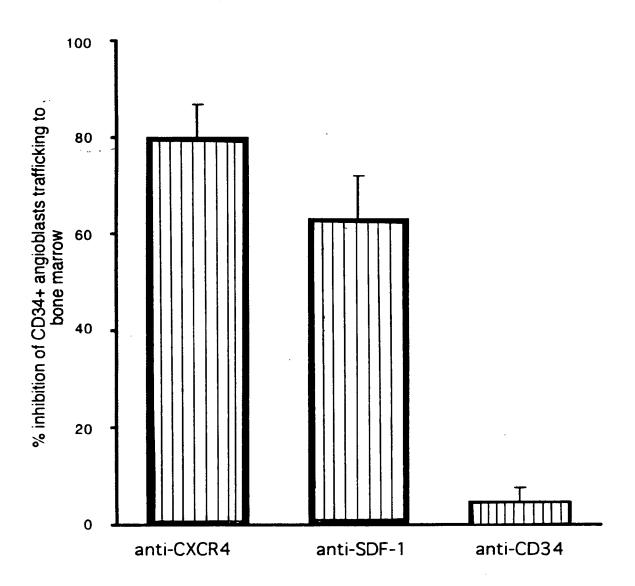
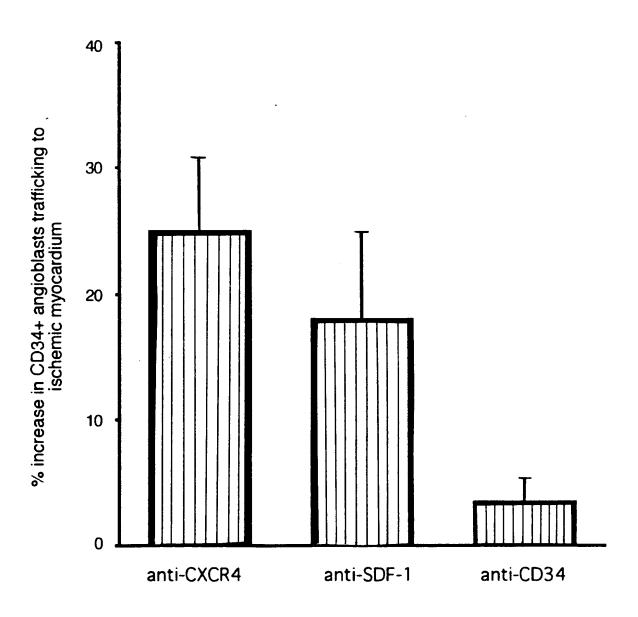
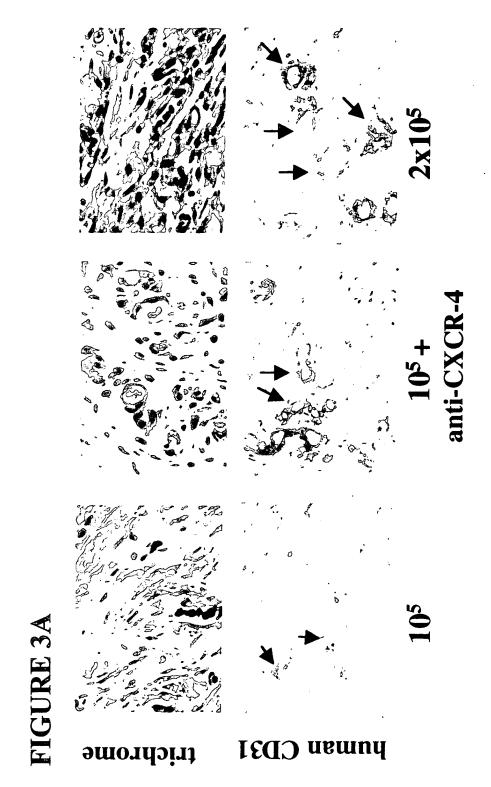
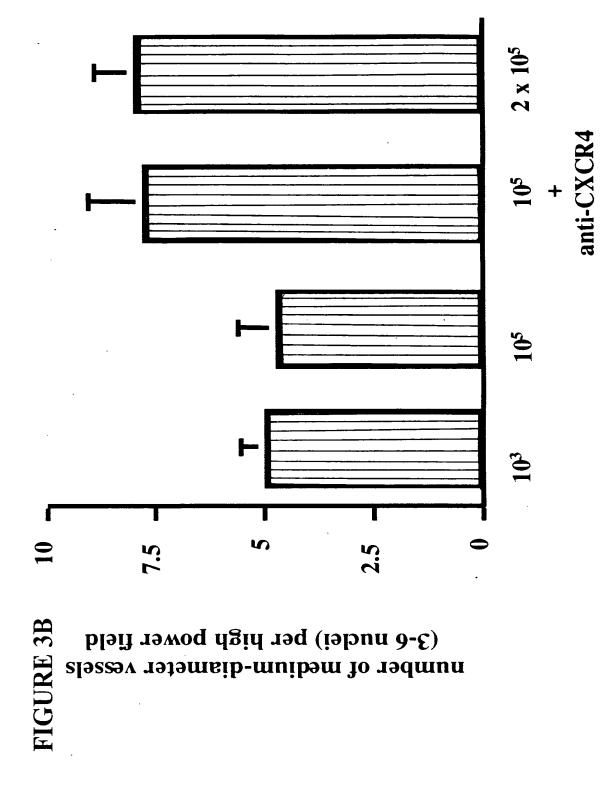


FIGURE 2C



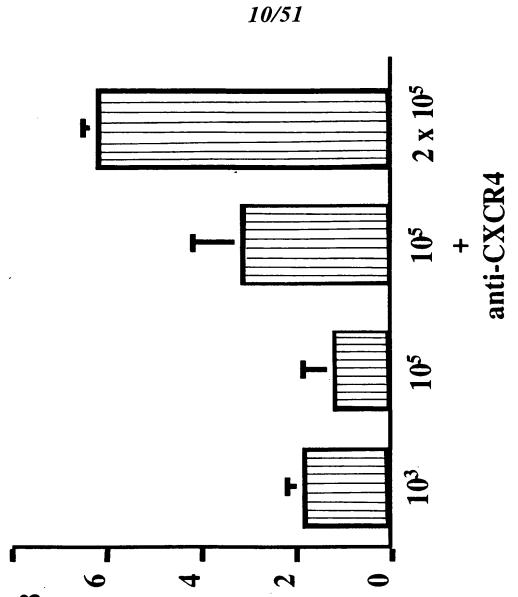


angioblast concentration (cells/mm³)

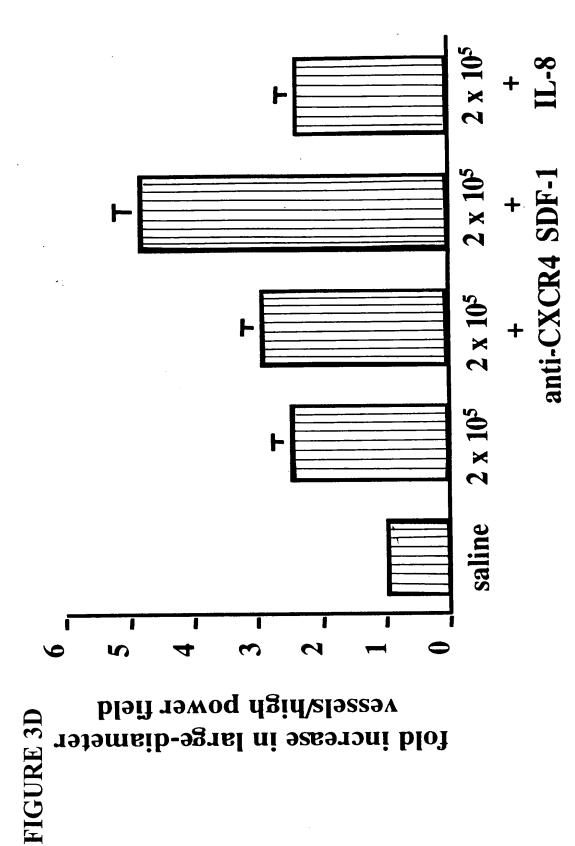


angioblast concentration (cells/mm³) and condition

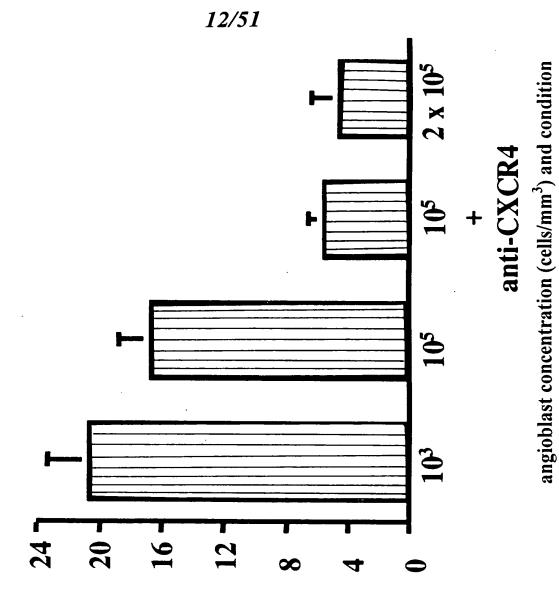
slager of large-diameter vessels (>6 nuclei) per high power field \$\infty\$



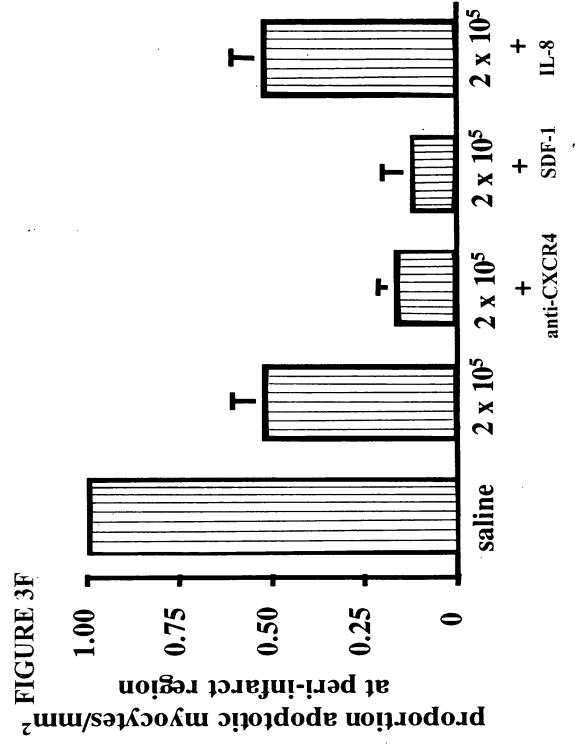
angioblast concentration (cells/mm³) and condition



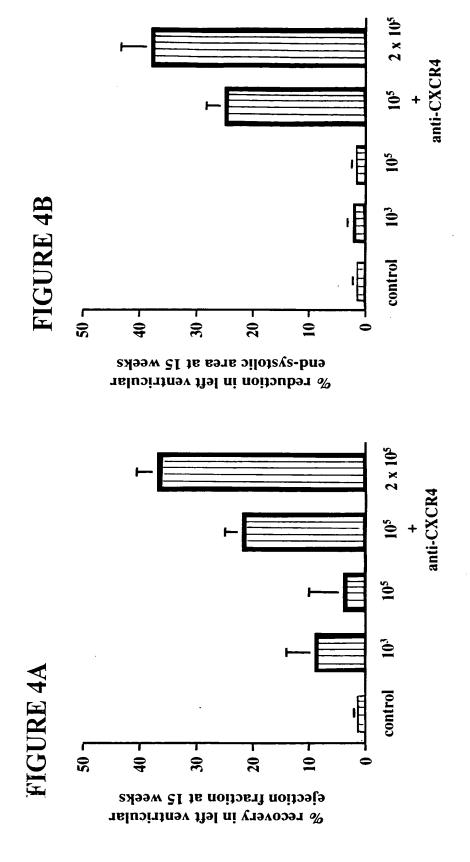
angioblast concentration (cells/mm³) and condition



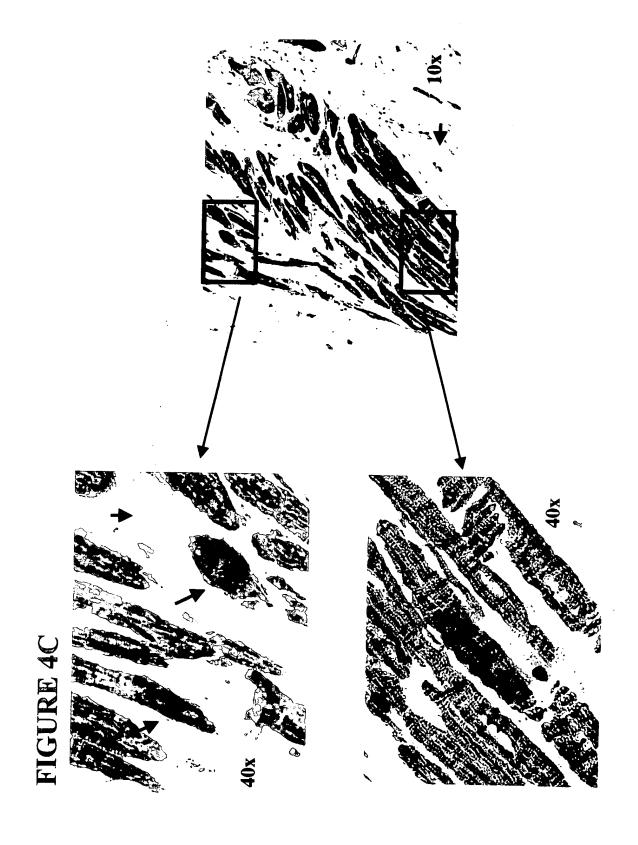
number of apoptotic myocytes/mm² at peri-infarct region



angioblast concentration (cells/mm³) and condition

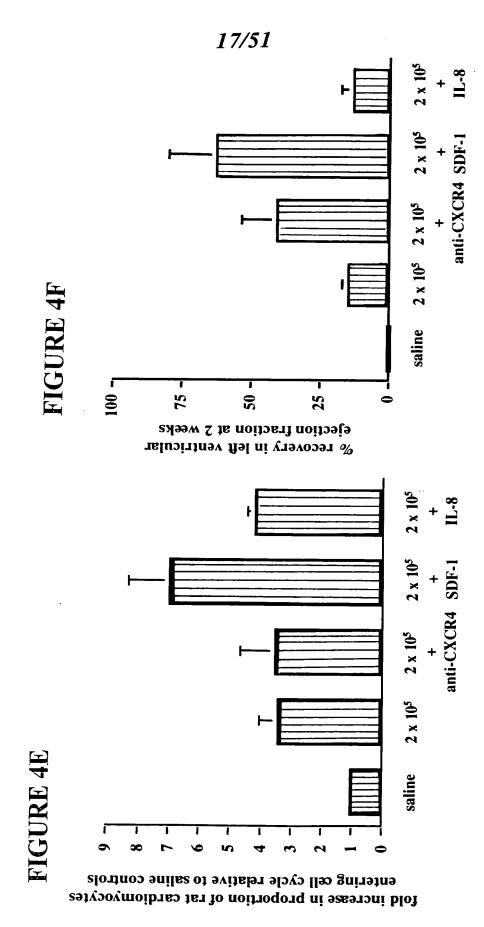


angioblast concentration (cells/mm³) and condition

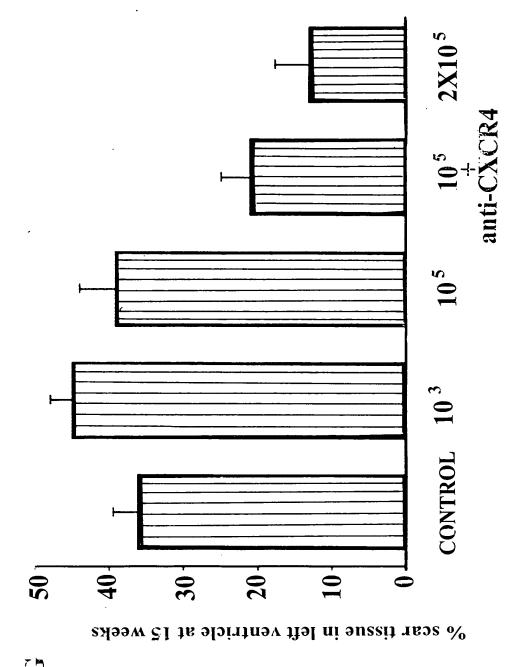


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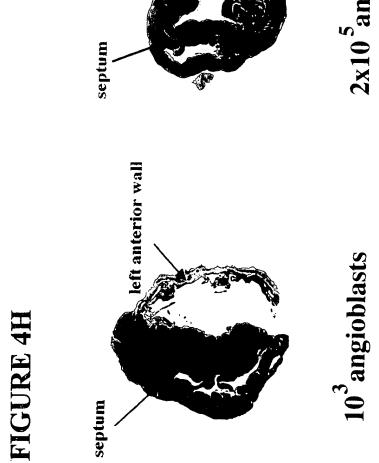
JURE 40



angioblast concentration (cells/mm³) and condition



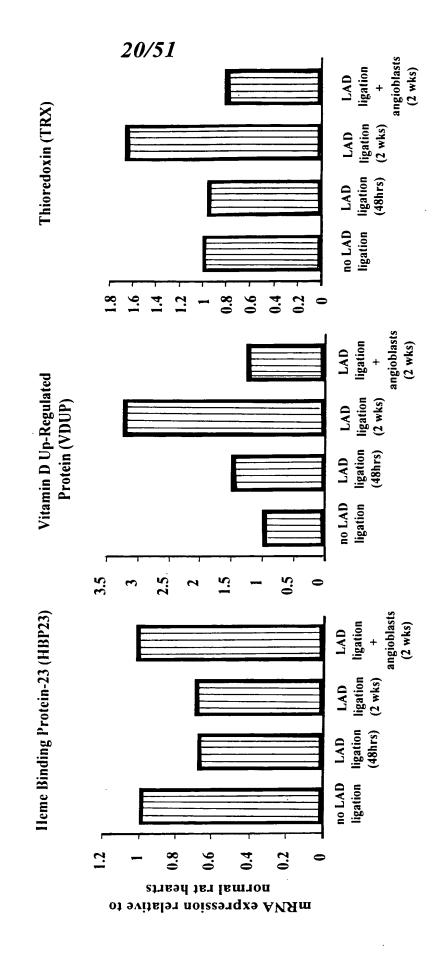
angioblast concentration (cells/mm³) and condition



left anterior wall

2x10 ⁵ angioblasts

FIGURE 5



21/51

FIGURE 6

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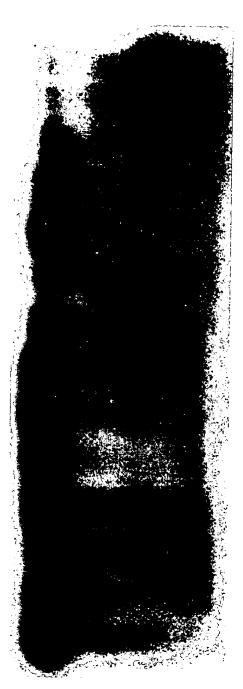
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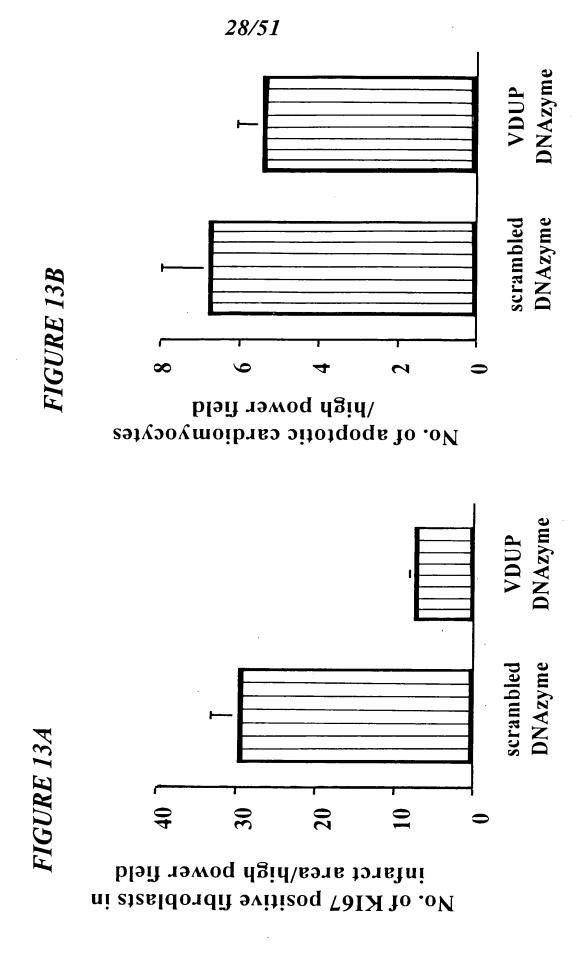
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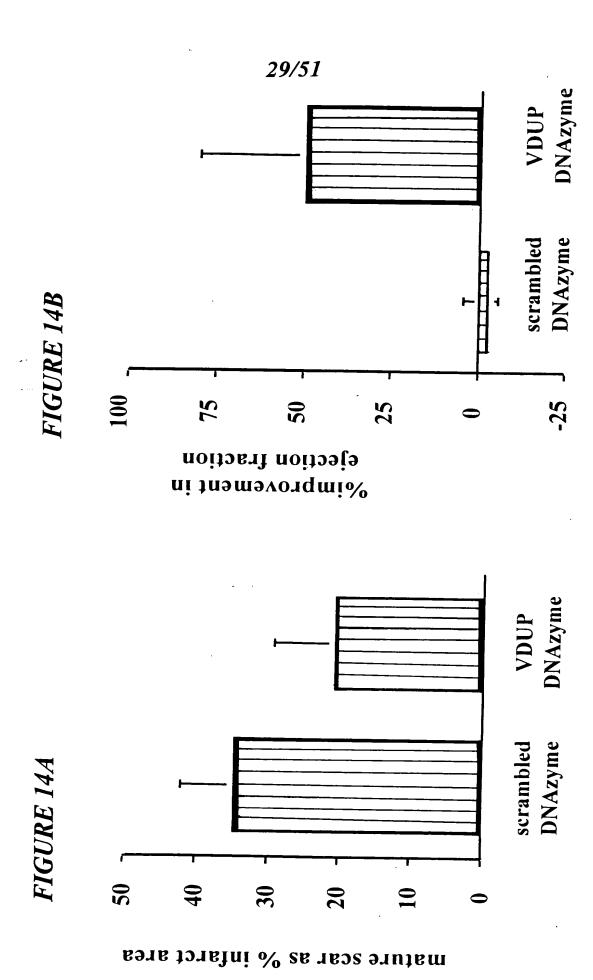
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FIGURE 12

	120	•
δμМ	09	
	20 60 120	
	w	
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0.5µM	20 60 120	
	09	
	20	
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	•	
0.05µM	120	
	60 120	
	S	
DNAzyme conc.	0	
	time (mins) 0	
	ţi	

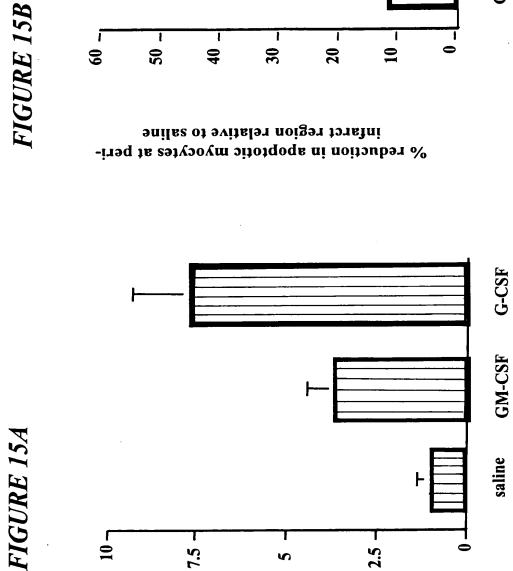






G-CSF

GM-CSF



30 _

50 -

9

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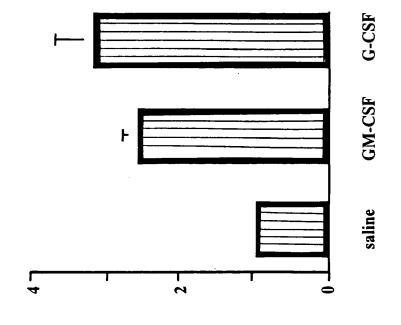
10 _

20 _

7.5 -2.5

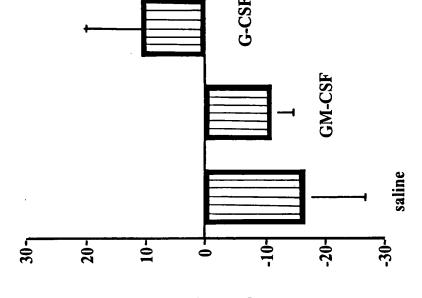
(>6 nuclei) per high power field number of large diameter vessels

entering cell cycle relative to saline controls fold increase in rat cardiomyocytes

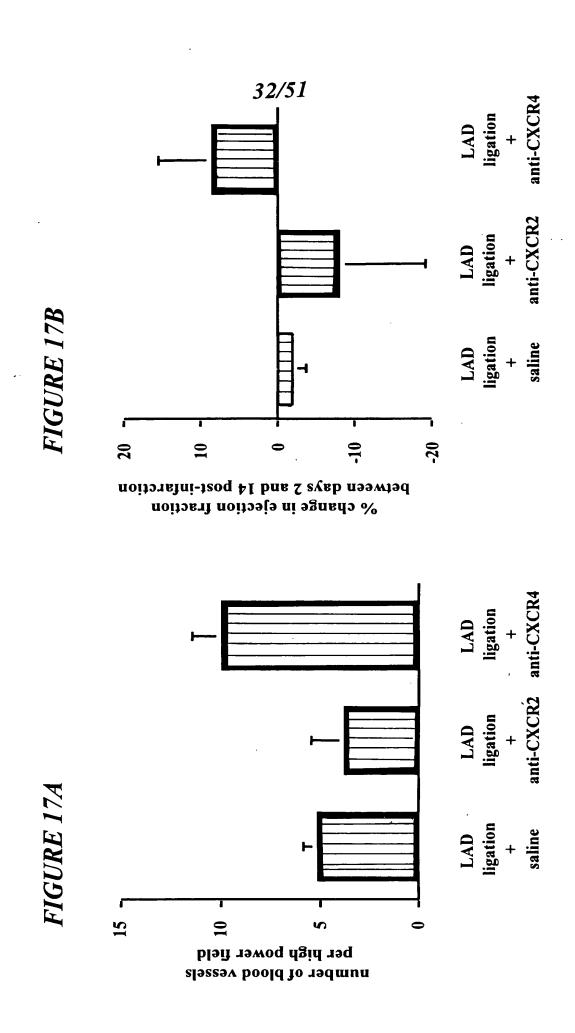


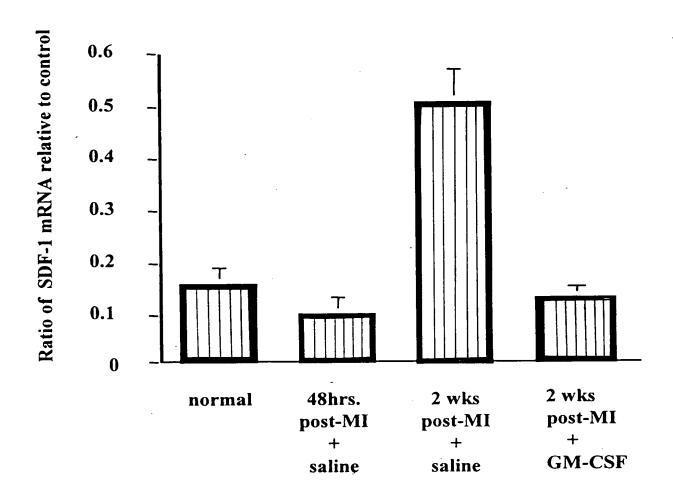
2 weeks post myocardial infarction % change in ejection fractionbetween 2 days and



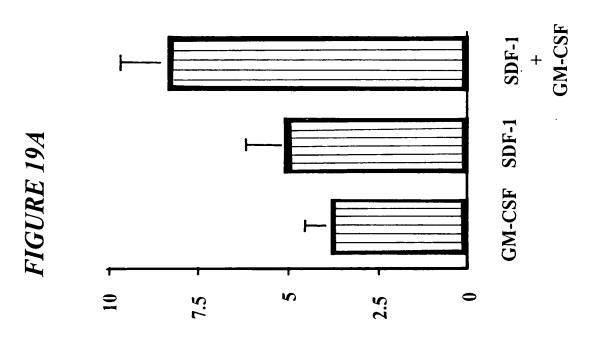


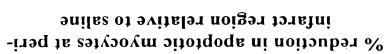
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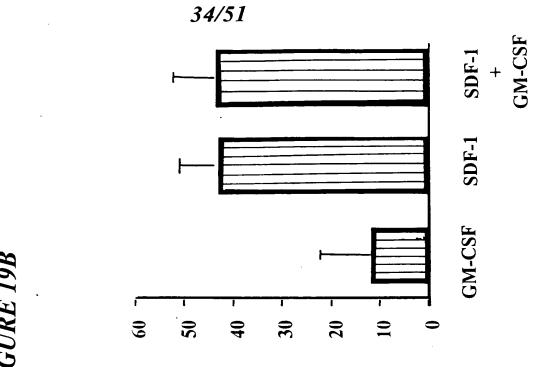




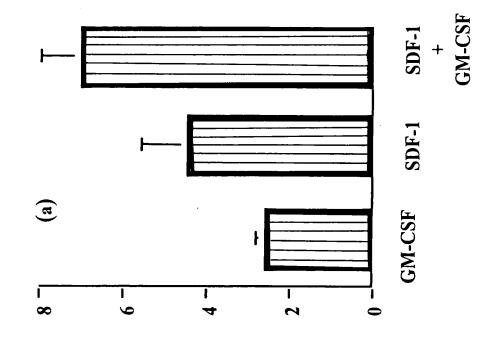
number of large diameter vessels (>6 nuclei) per high power field



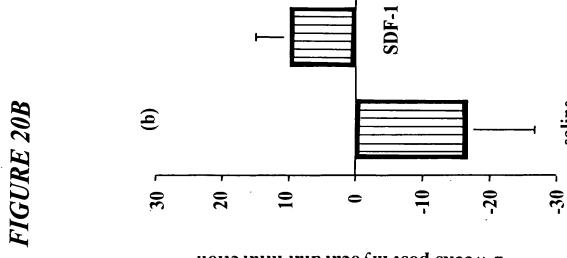




entering cell cycle relative to saline controls fold increase in rat cardiomyocytes

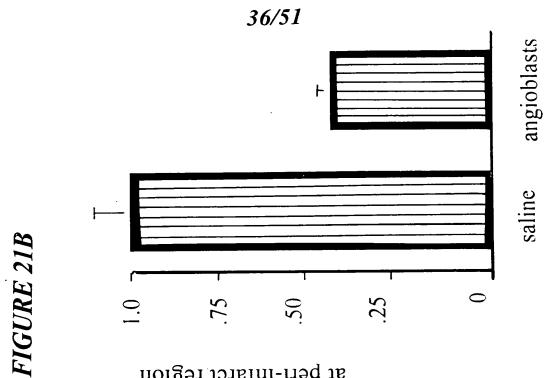


2 weeks post myocardial infarction 6 change in ejection fractionbetween 2 days and

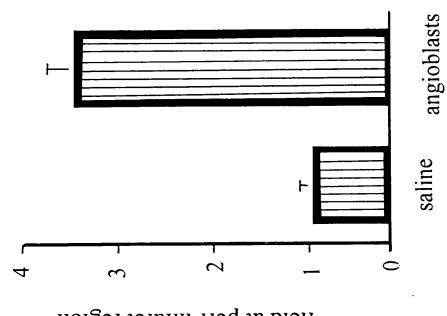


35/51





proportion of apoptotic myocytes at peri-infarct region

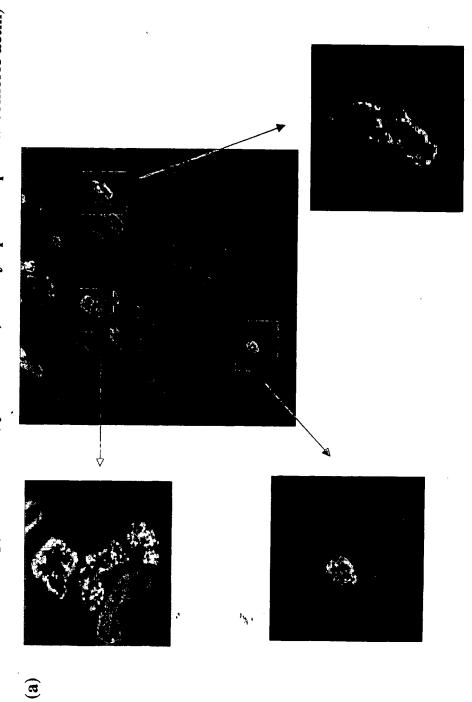


fold increase in vessels/high power field at peri-infarct region

FIGURE 22A

Early Proliferation/ Regeneration Of Immature Cardiomyocyte Lineage Progenitors Accompanying Neovascularization

(confocal microscopy: red nucleus, yellow Ki67, blue cytoplasm alpha-sarcomeric actin)

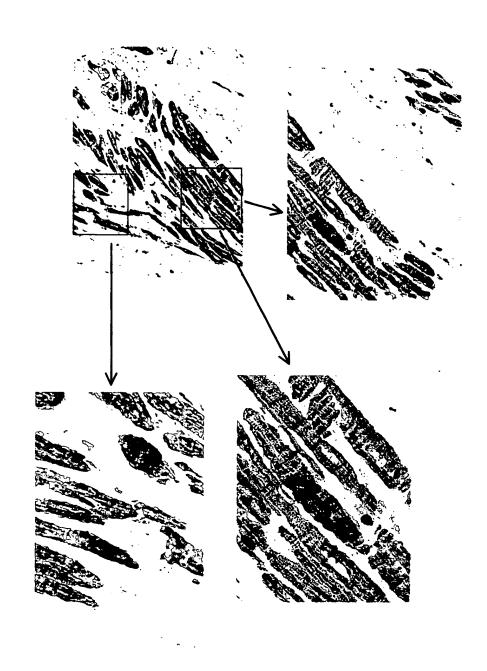


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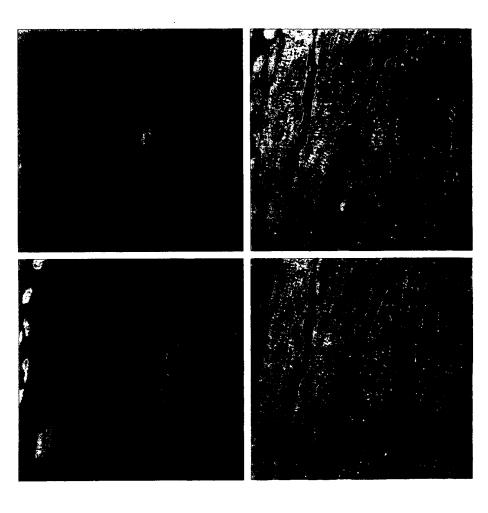
FIGURE 22B

(b) Later Differentiation And Regeneration Of Mature Cardiomyocytes Accompanying Neovascularization

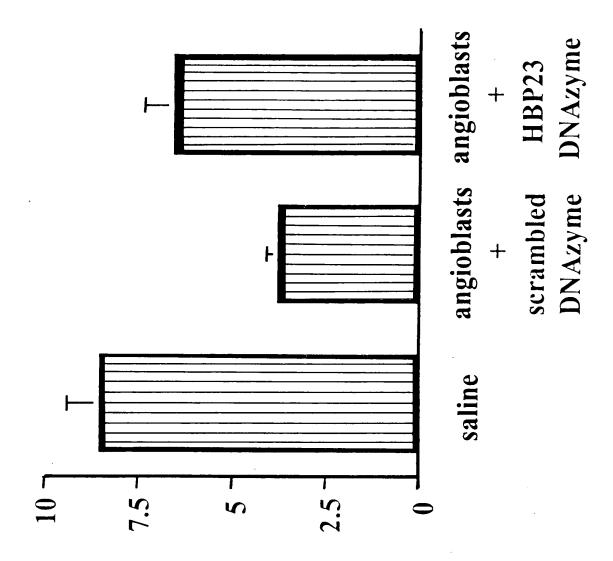
(immunohistochemistry: blue nucleus rat Ki67, brown cytoplasm troponin I)





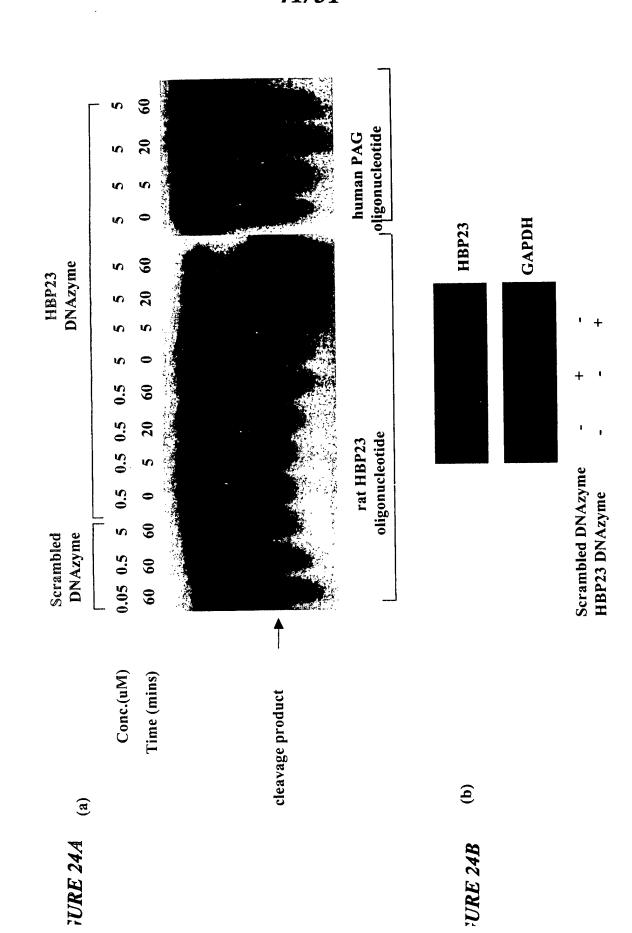


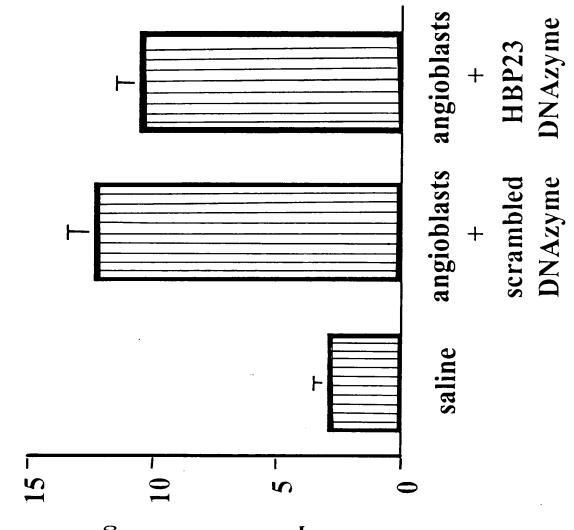
Confocal microscopy showing nuclear cycling (blue nucleus, green rat Ki67) of troponin I-positive mature cardiomyocyte (red cytoplasm)



number of apoptotic myocytes/high powered field at peri-infarct region

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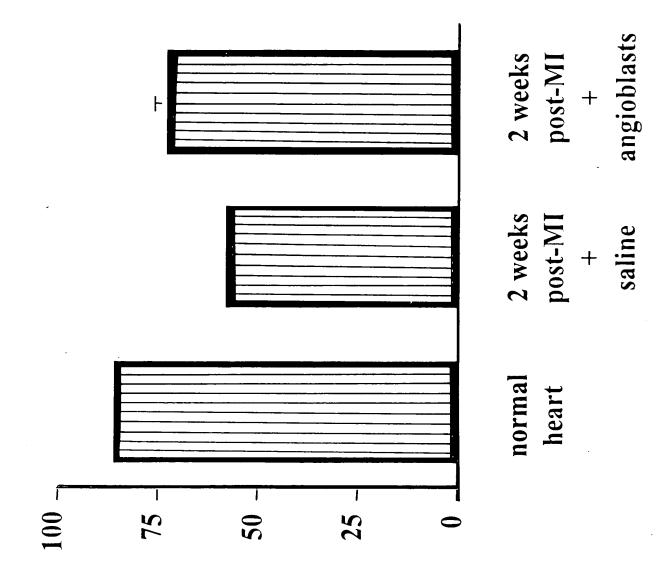




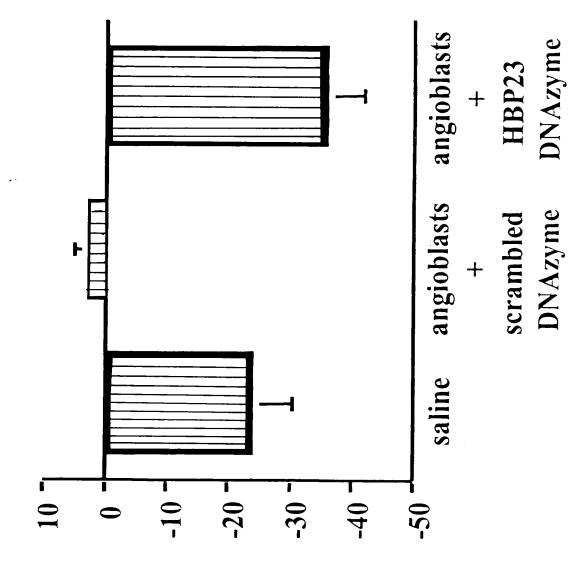
number of vessels/high power field at peri-infarct region

FIGURE 25A

relative expression of HBP23 mRNA



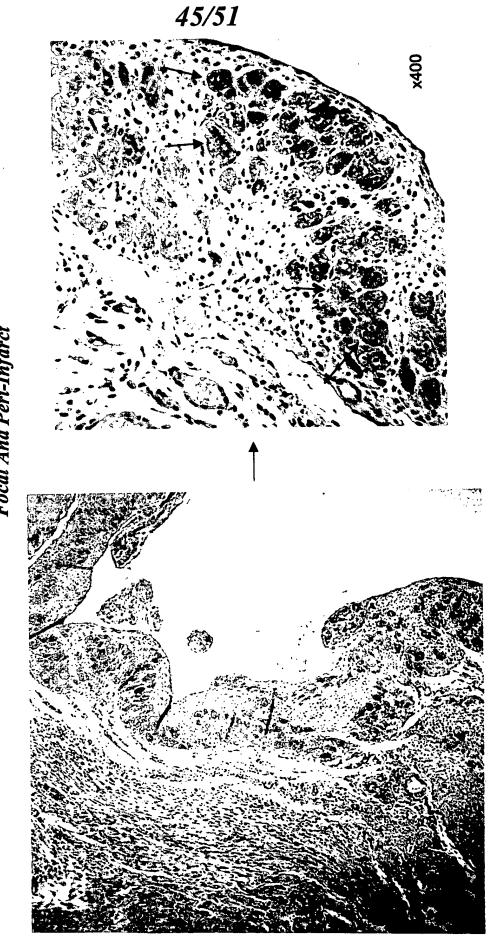
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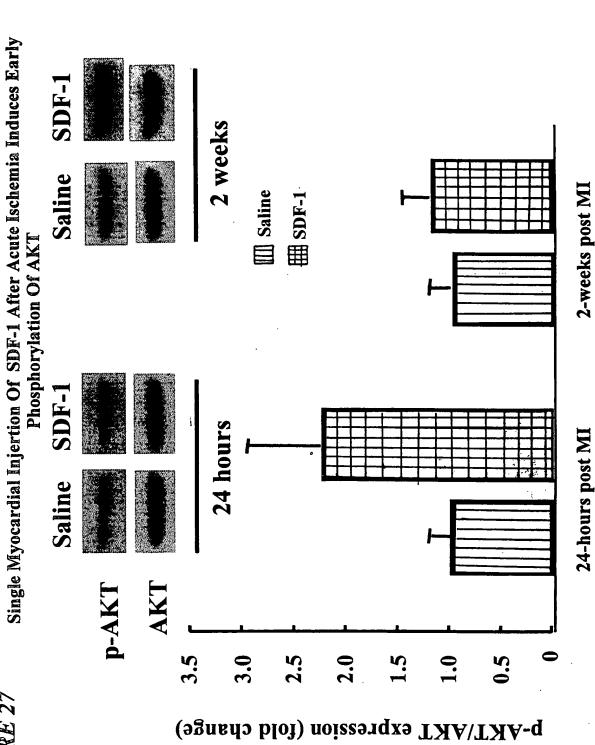


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FIGURE 26

Pattern Of CXCR4 Expression Following Acute Myocardial Ischemia Is Focal And Peri-Infarct





46/51

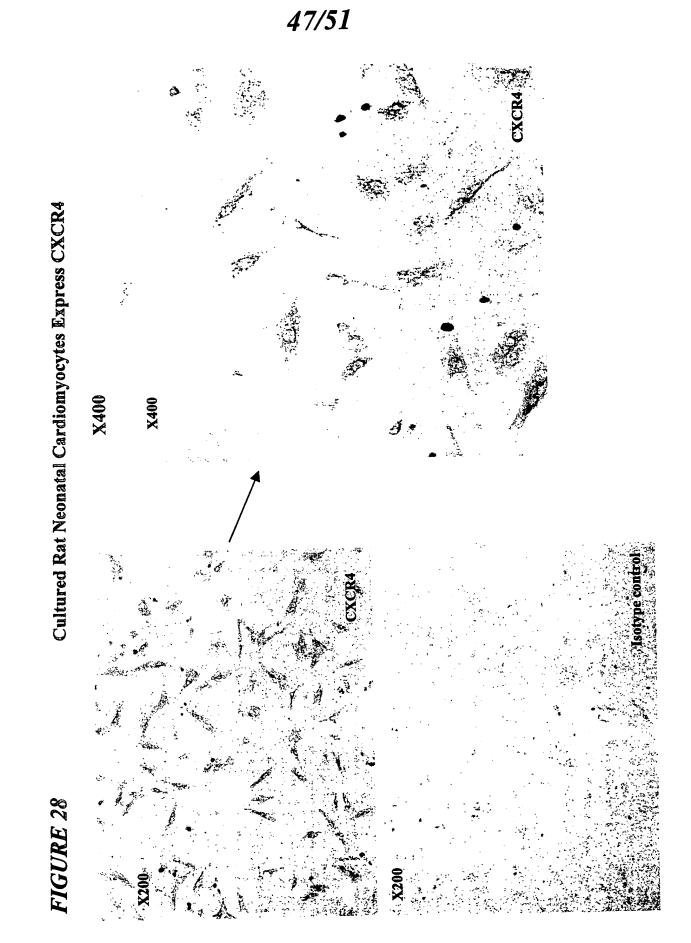
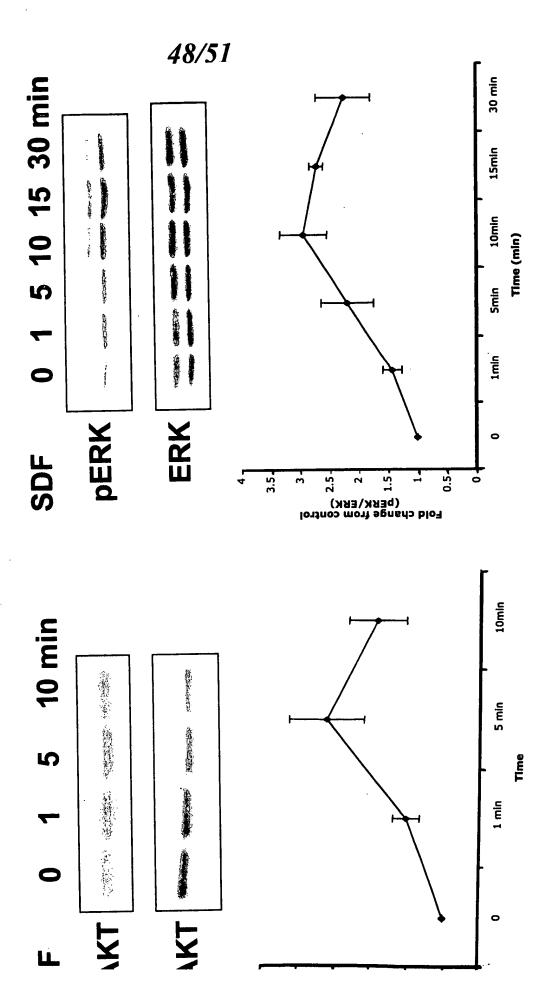
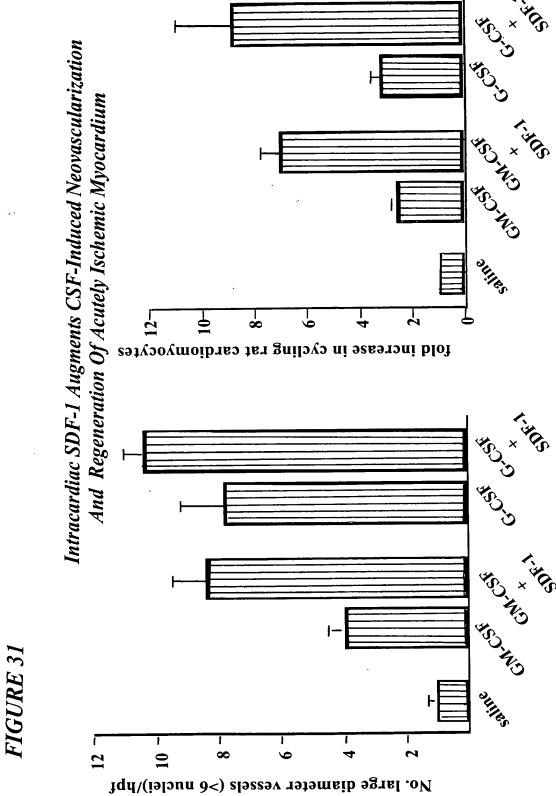
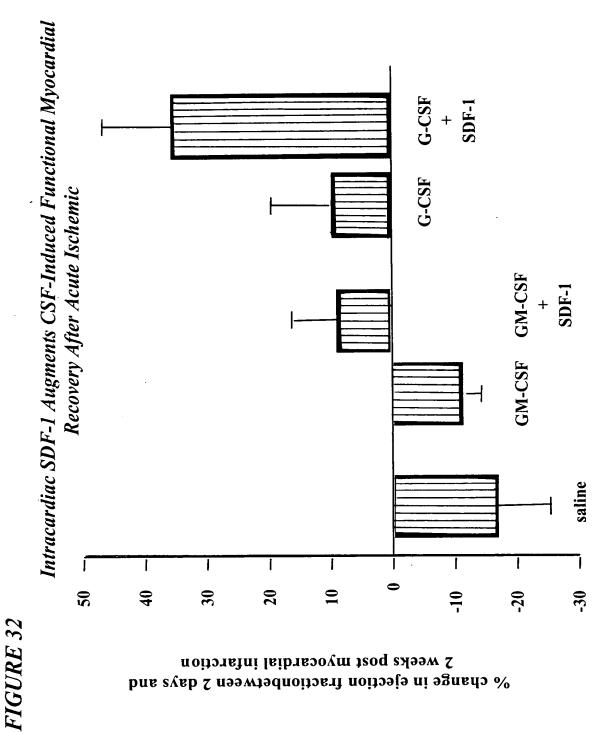


FIGURE 29

Effect of SDF (100nM) on pAKT/AKT and pERK/ERK Expression in Rat Neonatal Cardiac Myocytes







2 weeks post myocardial infarction % change in ejection fractionbetween 2 days and

1000 µM H2O2 10 µM SDF-1 .1 µM SDF-1 1 µM SDF-1 SDF-1 Protects Rat Neonatal Cardiomyocytes Against medium H202-Induced Apoptosis 40. 100. 80 -. 09 20 % Increase in viability 1000 10 µM SDF-1 500 medium 300 FIGURE 30 0 100 80 9 40 20 % Viability

51/51

[] H2O2 (µM)